

chromotek® HA-Trap Agarose

Product Code: ata

Product Information

Description: The ChromoTek HA-Trap Agarose consists of an anti-HA Nanobody/VHH, which is coupled to agarose beads. It can be used for the immunoprecipitation of HA-fusion proteins from cell extracts of various organisms such as humans, mice, dogs, yeast, and plants.

Applications: IP, Co-IP

Specificity/Target: Binds specifically to the HA-tag (sequence YPYDVPDYA) fused to a protein of interest at N-, C- or internal position. Please note that the affinity is highest for a C-terminal fusion. There is no cross-reactivity to other common peptide tags such as the His6-tag, FLAG-tag, Spot-Tag, V5-tag, Strep-tag, or C-tag (other tags not tested). Background binding to host cell proteins from a range of organisms such as human, mouse and dog cell lines or yeast and plants is low.

Binding capacity: 20 µg of recombinant HA-tagged protein (~30 kDa) per 25 µL bead slurry

Bead Size: 90 µm (cross-linked 4 % agarose beads)

Elution Buffer: 2x SDS-sample buffer (Lämmli)

Wash Buffer Compatibility: 1M NaCl, 5 mM DTT, 5 mM β-mercaptoethanol, 5 mM TCEP, 2% NP40, 2% Triton X-100, 0.1% SDS, 2-3 M Urea

Type: Nanobody

Class: Recombinant

Host: Alpaca

Shipment: Shipped at ambient temperature

Storage Buffer: 20 % ethanol

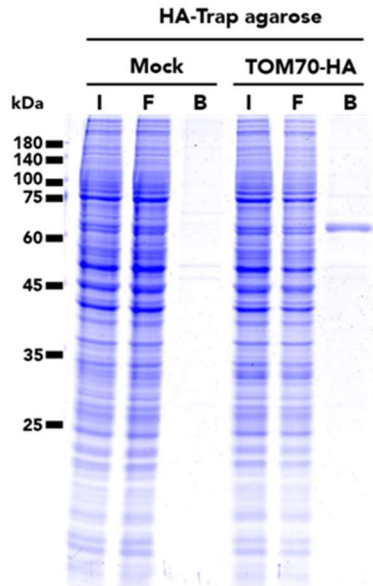
Storage Condition: Upon receipt store at +4°C. Do not freeze!

Stability: Stable for 1 year upon receipt

chromotek® HA-Trap Agarose

Product Code: ata

Selected Validation Data



Immunoprecipitation of TOM70-HA fusion protein from HEK293T cells using HA-Trap Agarose. IP was done using both un-transfected (mock) and transfected (TOM70-HA) cells. I: Input, F: flow-through, B: bound.

Suggested Buffer Compositions for IP

Buffer	Composition
Lysis Buffer	10 mM Tris/Cl pH 7.5, 150 mM NaCl, 0.5 mM EDTA, 0.5 % Nonidet™ P40 Substitute (adjust the pH at +4°C)
RIPA Buffer	10 mM Tris/Cl pH 7.5, 150 mM NaCl, 0.5 mM EDTA, 0.1 % SDS, 1 % Triton™ X-100, 1 % deoxycholate (adjust the pH at +4°C)
Dilution Buffer	10 mM Tris/Cl pH 7.5, 150 mM NaCl, 0.5 mM EDTA (adjust the pH at +4°C)
Wash Buffer	10 mM Tris/Cl pH 7.5, 150 mM NaCl, 0.05 % Nonidet™ P40 Substitute, 0.5 mM EDTA (adjust the pH at +4°C)
2x SDS-sample buffer	120 mM Tris/Cl pH 6.8, 20 % glycerol, 4 % SDS, 0.04 % bromophenol blue, 10 % β- mercaptoethanol
Acidic elution buffer	200 mM glycine pH 2.5 (adjust the pH at +4°C)
Neutralization buffer	1 M Tris pH 10.4 (adjust the pH at +4°C)
Peptide elution buffer	500 μM (0.64 mg/ml) HA-peptide (ap-1) reconstituted in PBS

Note: Use your equivalent cell lysis buffer for other cell types like yeast, plants, insects, bacteria.

Consider using a Wash buffer without detergent for Co-IP.

Note: Use Peptide elution buffer for elution under native conditions.

chromotek® HA-Trap Agarose

Product Code: ata

Product Sizes

Product	Product Code	Size
HA-Trap Agarose	ata-10	10 reactions
	ata-20	20 reactions
	ata-100	100 reactions

chromotek® HA-Trap Agarose

Product Code: ata

Protocol at a glance

General

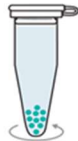
- Perform all steps at 4°C
- Use your preferred cell lysis buffer and cell lysis conditions

Cell Lysis



- Use 10^6 - 10^7 cells and 200 μ L Lysis buffer.
- Perform cell lysis and clear lysate
- Mix 200 μ L cleared lysate with 300 μ L dilution buffer.

Bead Equilibration



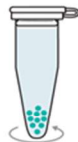
- Transfer 25 μ L bead slurry into a 1.5 mL tube
- Equilibrate beads 3x with 500 μ L dilution buffer

Protein binding



- Add 500 μ L diluted lysate to beads
- Rotate end-over-end for 1 hour at 4°C.

Washing



- Wash beads 3x with 500 μ L wash buffer
- Transfer beads to a new tube during the last washing step

Elution with SDS-sample buffer



- Resuspend beads in 80 μ L 2x SDS-sample buffer
- Boil beads for 5 min at 95°C
- Analyze the supernatant in SDS-PAGE/ Western Blot

chromotek® HA-Trap Agarose

Product Code: ata

Immunoprecipitation Protocol

Cell Material

The following protocol describes the preparation of a mammalian cell lysate.

For other type of cells, we recommend using 500 µg of cell extract and start the protocol with step Bead equilibration.

Mammalian Cell Lysis

Note: Harvesting of cells and cell lysis should be performed with ice-cold buffers. We strongly recommend adding protease inhibitors to the Lysis buffer to prevent degradation of your target protein and its binding partners.

For one immunoprecipitation reaction, we recommend using $\sim 10^6$ - 10^7 cells.

1. Choice of lysis buffer:
 - a. For cytoplasmic proteins, resuspend the cell pellet in 200 µL ice-cold Lysis buffer by pipetting up and down. Supplement Lysis buffer with protease inhibitor cocktail and 1 mM PMSF (not included).
 - b. For nuclear/chromatin proteins, resuspend cell pellet in 200 µL ice-cold RIPA buffer supplemented with DNase I (f.c. 75-150 Kunitz U/mL), $MgCl_2$ (f.c. 2.5 mM), protease inhibitor cocktail and PMSF (f.c. 1 mM) (not included).
2. Place the tube on ice for 30 min and extensively pipette the suspension every 10 min.
3. Centrifuge cell lysate at 17,000x g for 10 min at +4°C. Transfer cleared lysate (supernatant) to a pre-cooled tube and add 300 µL Dilution buffer supplemented with 1 mM PMSF and protease inhibitor cocktail (not included). If required, save 50 µL of diluted lysate for further analysis (input fraction).

Bead Equilibration

1. Resuspend the beads by gently pipetting them up and down or by inverting the tube. Do not vortex the beads!
2. Transfer 25 µL of bead slurry into a 1.5 mL reaction tube.
3. Add 500 µL ice-cold Dilution buffer.
4. Sediment the beads by centrifugation at 2,500x g for 5 min at +4°C. Discard the supernatant.

Note: Alternatively, ChromoTek Spin columns (sct-10; -20; -50) can be used to equilibrate the beads.

Protein Binding

1. Add diluted lysate to the equilibrated beads.
2. Rotate end-over-end for 1 hour at +4°C.

Washing

1. Sediment the beads by centrifugation at 2,500x g for 5 min at +4°C.

chromotek® HA-Trap Agarose

Product Code: ata

2. If required, save 50 µL of supernatant for further analysis (flow-through/non-bound fraction).
3. Discard remaining supernatant.
4. Resuspend beads in 500 µL Wash buffer.
5. Sediment the beads by centrifugation at 2,500x g for 5 min at +4°C. Discard remaining supernatant.
6. Repeat this step at least twice.
7. During the last washing step, transfer the beads to a new tube.

Optional: To increase stringency of the Wash buffer, test various salt concentrations e.g. 150-500 mM, and/or add a non-ionic detergent e.g. Triton™ X-100 (see Wash buffer compatibility table for maximal concentrations).

Note: Alternatively, ChromoTek Spin columns (sct-10; -20; -50) can be used to wash the beads.

Elution with 2x SDS-sample buffer (Laemli)

1. Remove the remaining supernatant.
2. Resuspend beads in 80 µL 2x SDS-sample buffer.
3. Boil beads for 5 min at +95°C to dissociate immunocomplexes from beads.
4. Sediment the beads by centrifugation at 2,500x g for 2 min at +4°C.
5. Analyze the supernatant in SDS-PAGE / Western Blot.

Note: For Western blot detection we recommend HA Tag Recombinant antibody (Proteintech 81290-1-RR) and Multi-rAb HRP-Goat Anti-Rabbit Recombinant Secondary Antibody (H+L) (Proteintech RGAR001).

Elution with Acidic Elution Buffer

1. Remove the remaining supernatant.
2. Add 50-100 µL Acidic elution buffer and constantly pipette up and down for 30-60 sec at +4°C or room temperature.
3. Sediment the beads by centrifugation at 2,500x g for 2 min at +4°C.
4. Transfer the supernatant to a new tube.
5. Immediately neutralize the eluate fraction with 5-10 µL Neutralization buffer.
6. Repeat this step at least once to increase elution efficiency.

Note: Elution at room temperature is more efficient than elution at +4°C. Prewarm buffers for elution at room temperature.

Elution with HA-peptide

1. Reconstitute 1 mg HA-peptide (ap-1) in 1550 µl PBS, which results in a final concentration of 500 µM (0.64 mg/ml). Vortex for 1 min to dissolve the powder.
2. Remove the remaining supernatant.
3. Add 80 µL HA-peptide (500 µM) and mix using a pipette.

chromotek® HA-Trap Agarose

Product Code: ata

4. Incubate at 25-37 °C for 5-10 min under regular pipetting to ensure thorough mixing.
5. Sediment the beads by centrifugation at 2,500x g for 2 min at +4°C.
6. Transfer the supernatant to a new tube.
7. Repeat this step at least once to increase elution efficiency.

Note: Elution will be most efficient for N-terminal and internal HA-tag fusions. For C-terminal HA-tag fusions, elute at 37 °C for up to 15 min.

Note: Alternatively, ChromoTek Spin columns (sct-10; -20; -50) can be used to separate the beads.

Related Products

Product	Code
HA-peptide	ap-1
HA-Trap Magnetic Agarose	atma
HA-Trap Magnetic Particles M-270	atd
HA-Trap Agarose Kit	atak
HA-Trap Magnetic Agarose Kit	atmak
HA-Trap Magnetic Particles M-270 Kit	atdk

Contact

Proteintech North America (HQ)

Proteintech Group, Inc.
5500 Pearl Street, Suite 400
Rosemont, IL 60018 USA

1-888-472-4522
proteintech@ptglab.com

Proteintech Europe

Transmission (6th Fl)
6 Atherton Street
M3 3GS, Manchester, UK

+44 161 839 3007
europe@ptglab.com

chromotek® HA-Trap Agarose

Product Code: ata

ChromoTek & Proteintech Germany

Fraunhoferstr. 1
82152, Planegg-Martinsreid
Germany

+49 89 124 148 850
germany@ptglab.com

Disclaimer

For research use only (RUO), not for diagnostic or therapeutic use.

ChromoTek® is a registered trademark of ChromoTek GmbH, part of Proteintech Group.